

Molecular Epidemiology of Cystic Echinococcosis: Genotypic Characterization in Humans and Different Livestock

Epidemiología Molecular de la Equinococosis Quística: Caracterización Genotípica en Humanos y Diferentes Animales

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SUMMARY: *Echinococcus Granulosus* (EG) is the major cause of cystic echinococcosis in humans and livestock in the world. In Chile is a zoonosis of great importance. The most frequently affected geographic areas are the Regions of Aysén, Los Rios, Los Lagos, Coquimbo and the Araucanía. Hence, it was discovered that in endemic areas of hydatidosis there could be several strains and genotypes of EG. In addition, there is evidence that some strains and genotypes are more infectious for human beings than others. This interesting phenomenon of the biology of EG has been studied using molecular biology techniques based on polymerase chain reaction (PCR) and DNA sequence analysis, which has made it possible to characterize the cestode species complex called EG sensu lato (s l) as being comprised of EG sensu stricto (s.s.) (Genotypes G1-G3), *E. equinus* (G4), *E. ortleppi* (G5) and *E. canadensis* (G6-G10), which present an important phenotypic variation detectable in characteristics of the biological cycle, specificity of the intermediate host, pattern of development, pathogenicity, antigenicity, transmission dynamics and, consequently, in the measures needed to control the disease. The aim of this manuscript is to describe the different genotypes of EG described in humans and different livestock host reported in the literature.

KEY WORDS: *Echinococcus granulosus*; Genotypes; cytochrome c oxidase subunit I; PCR-RAPD.

INTRODUCTION

Echinococcus granulosus (EG) is the major cause of cystic echinococcosis in humans and livestock in the world. In Chile is a zoonosis of great importance, as it constitutes a public health problem with strong socio-economic repercussions. The prevalence is 5 per 100.000 inhabitants nationally, and in the Region of the Araucanía it is 48 per 100.000 inhabitants (Fig. 1), particularly in rural areas where the population still incorrectly disposes of biological waste from animals slaughtered clandestinely (Serra *et al.*, 1995; Manterola *et al.*, 1997).

The high incidence and prevalence in these areas causes serious public health problems, including, it must be noted, the appearance of advanced stages of the disease with unusual forms of presentation and various types of complications of the lesions (Manterola *et al.*, 2003a; Manterola *et al.*, 2003b; Manterola *et al.*, 2004a; Manterola & Otzen 2017), which in turn increases the risks of morbidity

and mortality (Manterola *et al.*, 2004b) with the resulting rise in health costs.

In a study of the economic impact of human and animal echinococcosis in Chile, it was confirmed that i) in 2012, 767 patients were treated surgically for hydatidosis, ii) the annual costs derived from surgical treatment alone were estimated at USD 2.46 million, iii) when adding secondary costs due to sick leaves and loss of productivity, these costs increased to USD 3.13 million, and iv) considering human and animal costs, the annual economic burden of the disease was estimated at USD 14.35 million. The most frequently affected geographic areas are the Regions of Aysén, Los Rios, Los Lagos, Coquimbo and the Araucanía (Venegas *et al.*, 2014).

Therefore, this situation suggests a need to propose solutions that can be based on at least 4 possible perspectives:

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Fig. 1. Daughter's vesicles of a hydatid cyst of the liver in a human host.

education of the population (particularly those most susceptible to being infected), early diagnosis (especially in endemic areas), identification of risk factors (measure designed to recognize promptly those patients at greatest risk of disease-related morbidity and mortality), and the creation of a vaccine.

Hence, it was discovered that in endemic areas of hydatidosis there could be several strains and genotypes of EG (McManus & Smith, 1986; Bowles & McManus, 1993a), a fact that can impact various aspects of the epidemiology, pathology and control of the disease (Thompson & Lymbery, 1998). In addition, there is evidence that some strains and genotypes are more infectious for human beings than others (Eckert & Thompson, 1997). This interesting phenomenon of the biology of EG has been studied using molecular biology techniques based on polymerase chain reaction (PCR) and DNA sequence analysis, which has made it possible to characterize the cestode species complex called EG sensu lato (s.l.) as being comprised of EG sensu stricto (s.s.) (Genotypes G1-G3), *E. equinus* (G4), *E. ortleppi* (G5) and *E. canadensis* (G6-G10) (McManus & Rishi, 1989; Bowles *et al.*, 1992; Bowles *et al.*, 1993a; Bowles & McManus, 1993b; Bowles *et al.*, 1994; Siles-Lucas *et al.*, 1994; McManus *et al.*, 1995; Scott *et al.*, 1997), which present an important phenotypic variation detectable in characteristics of the biological cycle (Fig. 2), specificity of the intermediate host, pattern of development, pathogenicity, antigenicity, transmission dynamics and, consequently, in the measures needed to control the disease.

A great objective of these types of studies would be to develop vaccines against some specific genotypes. In this respect, some studies describe the difficulty of this initiative, given that the antigen of the protection vaccine (EG95), developed for the G1 genotype, is immunologically different in the G6 genotype; therefore, greater evidence is needed to



Fig. 2. Protoscolex obtained from hydatid sand of a human hydatid cyst. its intact membrane, calcareous corpuscles and chain hooks rostral appreciated.

determine whether the EG95 vaccine would be effective against the G6 or G7 genotypes, or if it would be necessary and possible to develop specific vaccines for each genotype (Alvarez Rojas *et al.*, 2014). The sequence of the full length of the cytochrome c oxidase subunit I (COI) (1609 bp) gene was analyzed in 37 EG isolates from different hosts and geographic regions in Australia, obtaining 7 haplotypes of EG s.s. not previously described, along with 5 haplotypes reported in other parts of the world, including the haplotype EG01, which is present in all the endemic regions. Such evidence points to further studies being required in the area (Alvarez Rojas *et al.*, 2016).

THEORETICAL FOUNDATIONS

EG is one of the smallest tapeworms in the Taeniidae family. This organism demonstrates a great variation in terms of specificity of the host, epidemiology, morphology, developmental biology, physiology, biochemistry and genetics (Thompson & Lymbery). Subspecific variants or strains have been identified, and a review of the taxonomy of the genus *Echinococcus* (Thompson *et al.*, 1995; Thompson, 2002) has been proposed. These subspecific variants are around 10 genotypes distinct from the parasite (G1 to G10) (Bowles *et al.*, 1992; Bowles *et al.*, 1994; Bowles *et al.*, 1993; Scott *et al.*; Lavikainen *et al.*, 2003), although the validity of the G9 genotype has been questioned because it could correspond to the G7 (Snábel *et al.*, 2000).

Several techniques have been applied to the direct study of the genetic variability of EG. Some working groups have focused their analysis on the regions of mitochondrial cytochrome regions of COI and NADH dehydrogenase 1 (ND1) of the parasite as genetic markers (Bowles *et al.*, 1993;

Bowles *et al.*, 1994; Bowles *et al.*, 1995; Zhang *et al.*, 1998a; Snábel *et al.*) based on the sequencing of the mitochondrial DNA and highlighting its importance in the differentiation and discrimination of organisms from the same family, as in the case of *Echinococcus* strains, which seem to be evolutionary units of high homogeneity (Bowles & McManus, 1993a; Bowles *et al.*, 1993b; Bowles *et al.*, 1992).

Previous studies into the identification and characterization of genotypes of EG have revealed the existence of nine genotypes, which have been linked to different animals, such as sheep, pigs, horses, camels, cattle, goats, etc. (Cuesta-Bandera *et al.*, 1988; Siles-Lucas *et al.*, 1993; Siles-Lucas *et al.*, 1996a; Siles-Lucas *et al.*, 1996b; Ponce-Gordo & Cuesta-Bandera, 1997a; Ponce-Gordo & Cuesta-Bandera, 1997b; Ponce-Gordo & Cuesta-Bandera, 1998). One of the criteria used in those studies was the band diagram analysis of random amplified polymorphic DNA (RAPD) (Olivier *et al.*, 1999). This method has proven to be fast and reliable, and has advantages over other techniques (larval morphology, in vitro development, isoenzyme standards and SDS-PAGE proteins); for example, RAPD can be used on infertile cysts because it requires small amounts of parasitic tissue, while it can also be applied to specimens that are fresh, frozen or fixed in alcohol. Nevertheless, the RAPD method has some drawbacks: it is very sensitive to changes in the reaction conditions (Olivier *et al.*), and it has low reproducibility and specificity due to the use of short primers (Ellsworth *et al.*, 1993; Micheli *et al.*, 1994; Valentini *et al.*, 1996; Ponce-Gordo & Cuesta-Bandera, 1997b; Ponce-Gordo & Cuesta-Bandera, 1998; Olivier *et al.*). Therefore, some groups have recommended the use of RAPD together with a PCR-restriction fragment length polymorphism (RFLP)-based analysis (PCR-RFLP) or DNA sequencing to obtain definitive results (Scott *et al.*). Preliminary results using these analytical methods were obtained by González *et al.* (2002), in the validation of an alternative method to discriminate EG genotypes in Spain, where, according to their results, there are only three (G1, G4 and G7). This method has an adequate internal and external validity, which makes it interesting to reproduce in other centers.

A PCR, followed by sequencing, on the mitochondrial genes COI and Nad1 is the gold standard test at laboratory level for addressing both the *Taenia* sp. determination as well the detection of *Echinococcus* sp. with subsequent species determination. The polymorphisms in these genes are so useful for typing (Roelfsema *et al.*, 2016).

In this light, EG is considered a tapeworm parasite of great medical and veterinary importance, as it is responsible for cystic echinococcosis, a condition that is among the most serious parasitic diseases in human beings,

representing 1 of the 17 neglected diseases prioritized by the World Health Organization (Laurimaa *et al.*, 2015).

AVAILABLE EVIDENCE

Bowles *et al.* (1994) of the Molecular Parasitology Unit of the Institute of Medical Research in Brisbane, Australia, studied the genetic characteristics of the “northern strain” of EG by applying a PCR analysis of the ITS1 nuclear region of the rDNA, finding that this variety presented ambiguities at position 18, which make it similar in structure to the G1 genotype (common in the domestic sheep), G2 (common in Tasmanian sheep) and G3 (common in buffalo). On the other hand, when they applied the mitochondrial ND1 sequence protocol, they observed a similarity with the G6 genotype (common in the camel) and the G7 (usual in pigs). The reactions observed support the notion that the “northern strain” of EG is a special genotype designated G8 (Bowles *et al.*, 1994).

Siles-Lucas *et al.* (1994), of the Department of Parasitology at the University of Madrid, Spain, used different molecular biology methods to examine isolates of EG from Switzerland and Spain, finding that the samples exhibited characteristic band patterns that confirmed that they belonged to horse and donkey isolates from Switzerland and Spain, cattle from Switzerland, sheep of Spain, and pigs from Switzerland and Spain, and goats from Spain (Siles-Lucas *et al.*).

Scott *et al.* (1997), of the Molecular Parasitology Unit of the Institute of Medical Research in Brisbane, Australia, analyzed EG isolates from pigs and humans in Poland, obtained using a fine needle puncture. The samples were processed by nuclear (ITS1) and mitochondrial sequencing (ND1) and then by application of restriction enzymes, observing that the Polish patients were not infected with the G1 genotype (associated with sheep and human infection), but rather a genotype similar to G7 (characteristic of pigs) was isolated in these patients, which represented a different genotype of EG that was called G9 (Scott *et al.*).

Zhang *et al.* (1998a), of the Molecular Parasitology Unit at the Australian Center for Tropical Diseases in Brisbane, Australia, examined 28 EG isolates from the Xinjiang Uygur region in China. The samples were obtained from sheep, cattle, camels and operated human hydatid cysts. After applying a DNA sequencing protocol to the mitochondrial cytochrome c oxidase subunit I and NADH dehydrogenase genes, they determined the existence of G6 (from camels) and G1 (from sheep and humans). The latter genotype was isolated exclusively in samples from the northern area of Xinjiang Uygur (Zhang *et al.*).

Zhang *et al.* (1998b), of the Molecular Parasitology Unit at the Australian Center for Tropical Diseases in Brisbane, Australia, studied 16 *Echinococcus* isolates from Iranian operated patients and from domestic animals in the geographic area of Tehran (including sheep, cattle, goats and camels). The samples were treated using a protocol to analyze the DNA nucleotides and to predict the sequence variations of the amino acids in the mitochondrial cytochrome c oxidase subunit 1 and NADH dehydrogenase genes. After applying a PCR and restriction enzymes, they found two different G6 (from dogs and camels) and G1 (from sheep, domestic animals and humans) genotypes (Zhang *et al.*).

Snabel. *et al.*, of the Institute of Parasitology at the Slovak Academy of Sciences in Kosice, Slovakia, determined the genotype of pig protoscolices in the Slovak territory, applying a mitochondrial (ND1) and nuclear (ITS1) sequence protocol, finding a high affinity of the samples with the G7 genotype (characterized by a double precipitation band pattern), which was the first report of the existence of the G7 genotype in Slovakia (Snabel *et al.*).

González *et al.*, of the National Microbiology Center in Madrid, Spain, verified 53 EG isolates in the central region of Spain (endemic for Echinococcosis). They extracted the complete genomic DNA, then amplified the DNA and this was automatically sequenced with a fluorescence base. The digested DNA samples were subjected to electrophoresis and hybridizations were done at 68 °C. The DNA sequences were analyzed by the program Primer Select Lasergen, and then amplified with PCR. The protocols made it possible to distinguish 2 genetic groups within the Spanish pig isolates: the G1 (sheep strain) and G7 (pig strain) genotypes. This observation can have a significant impact on public health care in Spain, since these pigs could infect human beings as has been reported in Eastern and Central Europe (González *et al.*).

Le *et al.* (2002), of the Unit of Molecular Parasitology at the International Center for Tropical Diseases in Brisbane, Australia, studied the complete mitochondrial genome of two genotypes of EG, G1 (sheep-dog strain) and G4 (horse-dog), which were compared with genomes from *E. multilocularis* and *Hymenolepis diminuta*, which had similar structures, lengths and composition (Le *et al.*).

Lavikainen *et al.* of the Department of Bacteriology and Immunology of the Haartman Institute at the University of Helsinki, Finland, analyzed 5 EG isolates from rodents in northeastern Finland, analyzing the DNA sequences of the mitochondrial cytochrome c oxidase subunit 1 and the NADH dehydrogenase, and the internal transcribed spacer ITS-1 of the ribosomal fragment. They observed that the

analyzed samples were similar to the G6 genotype; however, and according to the phylogenetic analysis, the isolates were also associated with the G7, G5 and G8 genotypes; as a result, it was considered that they were dealing with a new genotype, which they called "Fennoscandian cervid strain" and assigned it the G10 genotype (Lavikainen *et al.*).

Guarnera *et al.* (2004) of the Department of Parasitology at the National Institute of Infectious Diseases in Buenos Aires, Argentina, examined 41 patients with hepatic, pulmonary, splenic and multiorgan hydatidosis from the provinces of Neuquén and Tucuman, applying a protocol for sequencing mitochondrial COI. They found the G1 genotype (associated with common sheep) in 19 patients; the G2 genotype (related to sheep in Tasmania) in 6 patients, the G5 genotype (linked to cattle) in 1 patient, and the G6 genotype (associated with camels) in 15 patients. This finding is peculiar because the camel is not a common animal in Argentina (Guarnera *et al.*).

Daniel Mwambete *et al.* (2004), from the Department of Parasitology at the Complutense University of Madrid, Spain, verified cyst samples obtained from sheep, goats, cows, pigs and horses destroyed at local slaughterhouses in central Spain, to which specimens from human hydatid cysts and wild boars hunted in the central area of Spain were added. They worked with 248 isolates from hepatic and pulmonary cysts with a similar degree of fertility. They extracted the genomic DNA of the protoscolices or germinal membranes. They later applied PCR amplification reactions, finding 3 different genotypes: G1 (highly fertile and preference for sheep), G4 (no host preference and located exclusively in the liver of the infested animal), and G7 (preference for pigs, in which greater fertility was observed in hepatic than in pulmonary cysts) (Daniel-Mwambete *et al.*, 2004).

Mrad *et al.* (2005), of the Laboratory of Parasitology and Mycology at the Faculty of Pharmacy in Monastir, Tunisia, studied 372 cysts from 50 humans, 166 cattle, 153 sheep and 3 camels (all from Tunisia). After applying an analysis of the ITS sequence with PCR-RFLP, it was found that all the human, sheep and cattle samples corresponded to the G1 genotype and that the camel samples were related to G6 (Mrad *et al.*).

Boufana *et al.* (2008), of the Biomedical Sciences Research Institute at the University of Salford, UK, studied the performance of three PCR tests for the identification of the G1 genotype of EG in sheep, from canine tissue and fecal samples. The "Dinkel" and "Stefanic" primers were the most sensitive in the DNA detection of EG in dog feces and autopsies (73.7 % and 100 %, respectively). The "Abbasi" primers only detected 52.6 % of EG in the infected

dogs, but were more specific for the species, since the reaction crossed with *E. shiquicus* occurred in 90.9 % of the tissue samples and in 75 % of the fecal samples. The "Stefanic" primers were the least specific for EG (27.3 % in tissues and 25 % in feces). The "Dinkel" primers also showed crossed reactivity between species (63.6 % in tissues and 100 % in feces); however, they were more specific for the G1 strain of EG in sheep (Boufana *et al.*).

Manterola *et al.* (2008), of the Surgery Department at the Universidad de La Frontera, studied hydatid fluid and sand samples from patients operated on for hepatic and pulmonary hydatidosis in the regional hospital in Temuco, Chile, between 2004 and 2005. Two PCR systems were used: PCR Eg 9 (Fig. 3) and PCR Eg 16. The RSA I enzyme was used for RFLP.

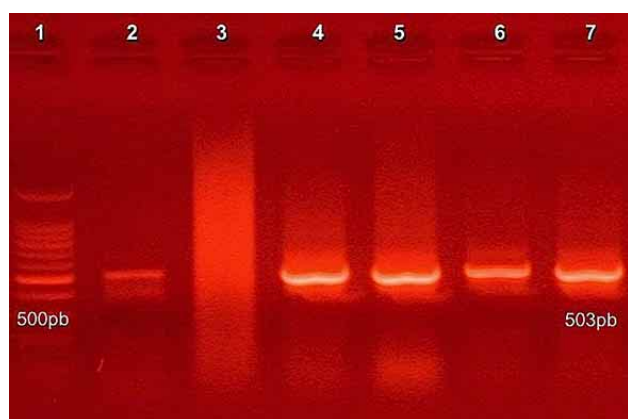


Fig. 3. EG9 PCR for DNA detection of *Echinococcus granulosus* in samples of human hydatid cyst. Agarose gel stained with 2 % ethidium bromide, shown in lane 1 marker 100 bp. In lanes 2, 4, 5, 6 and 7 positive samples. In lane 3, sample not amplified.

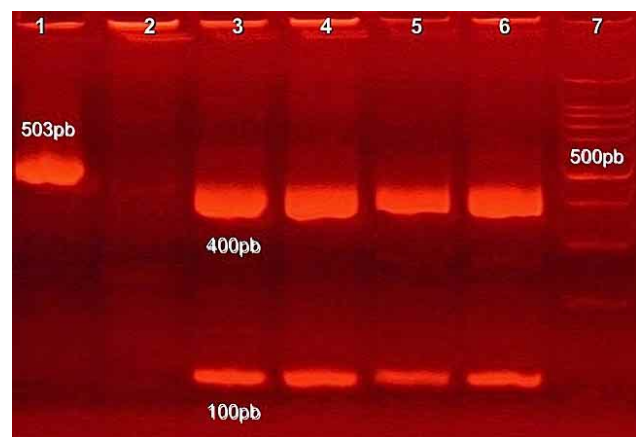


Fig. 4. EG9 PCR products digested with *RsaI* enzyme for DNA detection of *Echinococcus granulosus* in samples of human hydatid cyst. Polyacrylamide gel stained with 2 % ethidium bromide, shown in lane 1 a *RsaI* not cut. In lane 2 no amplified product. In lines 3, 4, 5 and 6 positive samples for enzymatic digestion. In line 7 marker 100 bp.

The genotype was confirmed using the sequence of a fragment of 366 base pairs from a mitochondrial gene (COI). The DNA of protoscolices from 24 samples (4 from pulmonary cysts and 20 from hepatic cysts) was analyzed. The fragment of 366 bp was amplified in 20 of the 24 samples (83.3 %). The enzymatic digestion (Fig. 4), revealed the presence of 3 possible genotypes: in 20 of 21 samples (95.2 %), a restriction was observed that corresponds to G1 or G7; in the remaining samples, it was determined that it could be G4 or G7. The final sequencing confirmed the presence of the G1 genotype in 19 samples and the G6 genotype for the remainder (G4 or G7 according to PCR-RFLP) (Manterola *et al.*).

Santivañez *et al.* (2008), of the Department of Microbiology at the Peruvian University Cayetano Heredia, Peru, conducted a molecular study by PCR using DNA from 21 hydatid cysts to determine the strains responsible for human infection in Peru. Amplification was applied using mitochondrial COI to 20 of 21 samples, revealing that all but one (95 %) belonged to the G1 strain in common sheep. The remaining samples belonged to the camel strain (G6) (Santivañez *et al.*).

Casulli *et al.* (2010), of the Department of Infectious and Parasitic at the Istituto Superiore di Sanità in Rome, Italy, performed a molecular analysis to identify EG genotypes causing human echinococcosis in the district of Turkana, Kenya (1993-1994). 71 cyst samples were taken using PAIR (puncture, aspiration, injection, reaspiration) procedures. The DNA of 59 isolates was amplified. Of these, 49 isolates (83 %) were identified as G1 and 10 (17 %) as G6 (Casulli *et al.*).

Soriano *et al.* (2010), of the Chair of Microbiology and Parasitology at the Universidad Nacional del Comahue (Neuquén, Argentina), studied the fertility, viability and molecular epidemiology of the hydatid cysts obtained from local goats (n = 23), pigs (n = 18) and sheep (n = 16) to identify possible reservoirs of *E. canadensis* (G6). Isolates from infected dogs (adult worms from 10 dogs) were also analyzed. They identified a total of 67 strains by DNA sequencing of mitochondrial cytochrome C oxidase subunit 1. They detected *E. canadensis* (G6) in 21 of 23 goat samples; *E. canadensis* (G7) in all the pig samples; EG sensu stricto (G3) in 1 sheep and G1 in 15 sheep, 2 goats and 9 dog samples. This was the first study to describe the existence of EG sensu stricto (G3) in South America and to verify that goats act as a reservoir of *E. canadensis* (G6) in Neuquén (Soriano *et al.*).

Nakao *et al.* (2010), of the Department of Parasitology at Asahikawa Medical College in Hokkaido, Japan, investigated the genetic polymorphisms of *Echinococcus* spp. in the eastern part of the Tibetan plateau and the Region of

Xinjiang, applying a DNA sequencing analysis to the mitochondrial COI and nuclear elongation factor-1 alpha. Thus, they obtained 68 strains of *E. granulosus sensu stricto* (S. S.) in Xinjiang. Additionally, 113 strains of *E. granulosus sensu lato*, 49 strains of *E. multilocularis* and 34 strains of *E. shiquicus* on the Tibetan plateau were isolated. Molecular identification using mitochondrial and nuclear markers was identical, suggesting a low frequency of introgressive hybridization (Nakao *et al.*).

Aaty *et al.* (2012), of the Faculty of Medicine at Ain Shams University, Abasia Square, Cairo, Egypt, analyzed isolates from 47 camels, 6 pigs and 31 humans (21 hepatic, 5 pulmonary and 5 multiorgan). The G6 genotype was verified in all the animal isolates (100 %) and in 30 of 31 (96.8 %) of the human isolates. This was done using the specific Hot-Start PCR followed by mitochondrial DNA 12S rRNA sequencing. The nucleotide sequence of the Egyptian G6 strain revealed 100 % homology with an Argentinian reference strain with 99 % homology with the G6 strain from Kenya. It was concluded that G6 is the predominant genotype in Egypt (Aaty *et al.*).

Andresiuk *et al.* (2013), from the Laboratory of Parasitic Zoonoses, FCEyN, UNMdP, Mar del Plata, Buenos Aires, Argentina, genetically characterized 42 isolates from cattle and 34 from sheep from the pampas in the province of Buenos Aires, Argentina. Using sequencing with mitochondrial COI and *nad1* genes, they verified that the predominant genotype was G1, with the exception of two identified as G2 and G5 (Andresiuk *et al.*).

Boufana *et al.* (2013) of the Cestode Zoonoses Research Group, University of Salford, UK, applied three PCR techniques based on the amplification of a fragment of the NADH dehydrogenase subunit 1 (ND1) in samples of *E. granulosus sensu stricto*, *E. shiquicus* and *E. multilocularis* obtained from parasitic tissue and fecal samples from dogs. A specificity of 100 % was verified when DNA derived from parasitic tissue was used, except for *E. shiquicus* primers, which barely detected DNA. The sensitivity of the three DNA detection techniques was between 2 and 10 gp. For the PCR-negative fecal samples, precipitation with ethanol was used to eliminate false negatives and increase sensitivity, as could be seen when the detection increased from 0 % to 89 % in the case of *E. shiquicus* coproDNA using samples from fox with an autopsy positive for echinococcosis (Boufana *et al.*).

Konyaev *et al.* (2013), of the Institute of Animal Ecology in Novosibirsk, Russia, studied a total of 75 samples of *Echinococcus* in European Russia and the Altai region between 2010 and 2012, finding the G6, G8 and G10 genotypes. In addition, 4 genotypes of *E. multilocularis* were identified (Konyaev *et al.*).

Espinoza *et al.* (2014), of the Agribusiness Faculty of the Universidad de La Frontera, analyzed samples of human and bovine hydatid cysts, for which they used segments of COI and NADH dehydrogenase (ND1). The phylogenetic analysis showed that most of the analyzed samples were G1; however, a sample of the cysts from a bovine lung (Q21), with the COI marker, was grouped in a node together with a sample belonging to the G3 genotype. In the phylogenetic tree obtained with the ND1 marker, this sample was grouped with the sequences of the G3, G2, and G4 genotypes. These results were significant as G2 and G3 had not been described previously in Chile (Espinoza *et al.*).

Boufana *et al.* (2014) of the Ecosystems and Environment Research Centre at the University of Salford, UK and other related centers, analyzed 174 *Echinococcus* isolates from human beings and ungulate intermediate hosts and adult tapeworms from dogs and jackals. They identified *E. granulosus sensu stricto*, from all the host species examined, *E. canadensis* (G6) in a camel, and for the first time the fertile cysts of *E. granulosus sensu lato* and *E. equinus* in equids (donkeys) and *E. granulosus sensu lato* in wild boars and goats. A considerable genetic variation for the *E. granulosus sensu stricto* COI (S. S.) sequences was observed. The pairwise fixation index (FST) for COI *E. granulosus sensu lato* sequences in donkeys was high and statistically significant compared to the populations of *E. granulosus sensu lato* in other intermediate hosts. A single haplotype (EqTu01) was identified by the nucleotide sequences of equine *Echinococcus* (Boufana *et al.*).

Alvarez Rojas *et al.* (2014), of the Faculty of Veterinary Sciences at the University of Melbourne, Australia and other related centers, suggested that the allocation of samples to G numbers is becoming increasingly difficult, as there is greater variability than previously known in the G1-3 (*E. granulosus sensu stricto*), *E. equinus* (G4), *E. ortleppi* (G5) and *E. canadensis* (G6-7, G-8, G10) genotypes. The G1 genotype is responsible for most of the human cystic echinococcosis (88.4 % worldwide). The G6 and G7 genotypes (closely related) cause a significant number of human infections (11.1 %). G5, G8 and G10, however, have been identified in human samples, but with less frequency. Cases of human infection by G4 have not been described. Finally, they proposed that the biological differences between species and genotypes have the potential to affect the transmission dynamics of the parasite (Alvarez Rojas *et al.*).

Babazadeh *et al.* (2015), from the Department of Clinical Sciences at the School of Veterinary Medicine at the University of Shiraz, Iran, analyzed DNA samples from protoscolices extracted from hepatic hydatid cysts in a two-year-old male water buffalo, slaughtered in the province of Khuzestan. A comparative analysis was made of COI

mitochondrial gene sequences with which phylogenetic molecular analyses were conducted. The existence of a new microvariant of the G3 genotype was found in Iran, with genetic differences from the G3 reference sequence (M84663). The difference is a transition mutation from adenine to guanine at position 214 (A214G), which is a substitution of threonine (ACT) for alanine (GCT). This contributes to knowledge of the heterogeneity and distribution of the G3 genotype for EG in the world (Babazadeh *et al.*).

Laurimaa *et al.*, from the Zoology Department of the Institute of Ecology and Earth Sciences at the University of Tartu, Estonia, studied fecal sample from 181 stray dogs in an urban area of Estonia in 2012-13, reporting positivity for EG in 2.2 % of the cases, with sequencing for the G1 genotype (positivity was not verified for *E. multilocularis*).

Boufana *et al.* (2015) of the Ecosystems and Environment Research Centre at the University of Salford, UK and other related centers, studied hydatid cysts isolated in horses, cysts recovered from mammals in captivity, and coproDNA from hunting dogs and farm dogs, identifying *E. equino* from the horse samples, EG (S. S.) in cattle and sheep samples and from farm dog feces. A variation in the haplotypes of EG (S. S.) was detected in the COI sequences (Boufana *et al.*).

Cucher *et al.* (2016), Research Institute for Medical Microbiology and Parasitology at the University of Buenos Aires, Argentina and other related centers, performed a systematic review of the literature, in which they found information geared toward South America in Argentina, Brazil, Chile, Peru and Uruguay, reporting sequencing data from 1534 cases of EG s. s. (G1). Similarly, the analysis of the cases all over the world showed that 72.9 % of the cases in humans are caused by EG s. s. (G1), and 12.2 % and 9.6 % by *E. canadensis* G6 and G7, respectively (Cucher *et al.*).

Spotin *et al.* (2016) of the Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran and other related centers, sequenced 79 isolates from different host species covering human, dog, camel, goat, sheep and cattle as of various geographical sub-populations of Iran (Northwestern, Northern, and Southeastern), and 36 sequences of other geographical populations (Western, Southeastern and Central Iran). The confirmed isolates were grouped as G1 genotype (n=92), G6 genotype (n=14), G3 genotype (n=8) and G2 genotype (n=1). They also verified that neutrality indices of the COI were shown negative values in Western-Northwestern, Northern and Southeastern populations which indicating significant divergence from neutrality and positive but not significant in Central isolates (Spotin *et al.*).

Eslami *et al.* (2016), from the Department of Parasitology, Faculty of Veterinary Medicine, University of Tehran, Iran, determined the characteristics of genotype and phenotype of EG derived from wild sheep comparing them with the strains of EG s.s. (sheep-dog) and EG camel strain (camel-dog) in Iran. They verified that the lengths of ITS2 and COI sequences were 378 and 857 bp, respectively, for all the sequenced samples. The amplified DNA sequences from both ribosomal and mitochondrial genes were 99 % and 98 %, respectively to that of the ovine strain in the GenBank database. So they conclude that the morpho-molecular features and characteristics of EG in the Iranian wild sheep are the same as those of the sheep-dog EG s.s. Strain (Eslami *et al.*).

Zait *et al.* (2016) of the Parasitology and Mycology department, Mustapha University Hospital, Algeria and other related centers, studied 54 isolates from humans admitted in surgical units from Mustapha Hospital and 16 cysts from livestock animals gathered in two geographically distinct slaughterhouses (Tiaret and Tamanrasset). They performed molecular characterization using sequencing of two mitochondrial genes, COI and NADH dehydrogenase subunit I (NDI). The study confirmed EG G1 as the main genotype (90.7 %) followed by G3 (7.4 %) and *E. canadensis* G6 (1.8 %). On the other hand, all cysts from sheep, cattle, and goat were identified as EG G1 and the two cysts originating from dromedary as *E. canadensis* G6. Twenty concatenated haplotypes (COI + NDI) were characterized. Among EG s.s., one haplotype (HL1) was highly predominant in both humans and animals cysts (71.6 %) (Zait *et al.*).

Sharbatkhori *et al.* (2016), of the Infectious Diseases Research Center, Golestan University of Medical Sciences, Gorgan, Iran and other related centers, studied 74 hydatid cysts isolated from livestock (18 from sheep, 40 from cattle, 9 from camels, 2 from buffaloes and 1 from a goat) and humans (4 isolates) in the Golestan province, northern Iran, southeast of the Caspian sea, using partial sequencing data of COI and NADH dehydrogenase I mitochondrial genes. Then, isolates were analysed by PCR amplification and sequencing. The results observed were G1 (78.3 %), G2 (2.7 %), G3 (15 %) and G6 (4 %) (Sharbatkhori *et al.*).

Oudni-Mrad *et al.* (2016) of the Laboratory of Medical and Molecular Parasitology-Mycology, Faculty of Pharmacy, University of Monastir, Tunisia and other related centers, studied G1 genotype of EG s.s. genetic relationship between lung and hepatic localization in ovine, bovine and human echinococcosis. They used allozyme variation and single strand conformation polymorphism for genetic differentiation and they found that the causes of genetic differentiation were the host species and the localization (lung or liver). Thus it can be assumed that reticulated genetic relationship between liver or

lung human isolates and isolates from bovine lung, is indicative of recombination or lateral genetic transfer (Oudni-Mrad *et al.*).

Roiniotti *et al.* (2016) Department of Anatomy and Physiology of Farm Animals, Faculty of Animal Science and Aquaculture, Agricultural University of Athens and other related centers, conducted a genetic analysis of 82 EG isolates from ruminants in Greece. The analysis relied on a PCR assay targeting COI, followed by bidirectional sequence analysis of the amplification product. 97.6 % isolates were allocated to EG s.s. (G1-G3) and were classified in 13 distinct haplotypes (9 common and 4 novel) with 12 polymorphic sites. All nucleotide changes proved to be transitions (A↔G or C↔T). Two fertile hydatid cysts of sheep origin of the Peloponnese were identified as *E. canadensis* (G7 genotype) (Roiniotti *et al.*).

The previously mentioned studies reveal the great variability of genotypes found in the different hosts and different areas of the world.

But only two genotyping experiences in our country (both in La Araucanía), with a limited number of samples (Manterola *et al.*, 2008; Espinoza *et al.*, 2014) is undoubtedly insufficient to have proper information regarding the great variability described in other latitudes, particularly when different techniques for genotype identification have been used.. This is the reason we believe that this proposal will be an important contribution to the knowledge of this disease in our country, from which valuable information can be obtained to advance in the development of preventive measures to reduce the high prevalence rates recorded in both animals and humans (for example, the development of specific vaccines against the most prevalent genotypes

MANTEROLA, C. & OTZEN, T. Epidemiología molecular de la equinococosis quística: Caracterización genotípica en humanos y diferentes animales. *Int. J. Morphol.*, 34(4):1472-1481, 2016.

RESUMEN: *Echinococcus granulosus* (EG) es la principal causa de equinococosis quística en humanos y ganado en el mundo. En Chile hay una zoonosis de gran importancia. Las zonas geográficas más afectadas son las Regiones de Aysén, Los Ríos, Los Lagos, Coquimbo y la Araucanía. Por lo tanto, se descubrió que en áreas endémicas de hidatidosis podría haber varias cepas y genotipos de EG. Además, hay pruebas de que algunas cepas y genotipos son más infecciosos para los seres humanos que otros. Este interesante fenómeno de la biología del EG ha sido estudiado utilizando técnicas de biología molecular basadas en la reacción en cadena de la polimerasa (PCR) y análisis de secuencias de ADN, lo que ha permitido caracterizar el complejo de cestode llamado EG sensu lato (sl) EG (G3) y *E. canadensis* (G6-G10), que presentan una importante variación fenotípica detectable en las características del ciclo biológico, especificidad del huésped intermedio, patrón de desarrollo, patogenicidad, antigenicidad, dinámica de transmisión y, por consiguiente, en las medidas necesarias para el control de la enfermedad. El objetivo de este manuscrito fue describir los diferentes genotipos de EG descritos en humanos y diferentes animales de ganado reportados en la literatura.

PALABRAS CLAVE: *Echinococcus granulosus*; Genotipos; Citocromo c oxidasa subunidad I; PCR-RAPD.

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